

ANTHOCYANIN ISOLATION METHOD OF RED DRAGON FRUIT (Hylocereus polyrhizus): A REVIEW

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ABSTRACT

Background: The body needs antioxidants to reduce the number of free radicals in the body. The anthocyanin compounds in red dragon fruit can be a source of antioxidants to counteract free radicals. Many types of solvents can be used to extract anthocyanins, such as water, methanol, or ethanol. The extracts produced from this process usually contain solvents that can reduce the anthocyanin content in the extract. Objective: Researchers believe that a literature review is needed to find out the factors that affect the isolation of anthocyanin compounds from dragon fruit skin, such as extraction methods, types of solvents, temperatures, and pH. Methods: The methods used in this study were obtained from Google, Google Cendekia, and Pubmed using keywords such as anthocyanin isolation and red dragon fruit. The article used was selected in the range of 10 years, from 2014 to 2024. The inclusion criteria in the search for literature are the articles that explain the anthocyanin isolation method of red dragon fruit and articles are available in Indonesian or English, while the exclusion criteria in this study are articles that are an article review. Results: The methods of maceration, Ultrasonic Assisted Extraction, Microwave Assisted Extraction, and Microwave Assisted Hydro Distillation can be used to isolate anthocyanin compounds using solvents such as citric acid, ethanol, and acetic acid. Conclusion: The ultrasonic assistance method is known to be able to attract the most anthocyanin compounds from red dragon fruit compared to other methods, which is 29,640 mg/5 g. The solvent with the acid environment is more suitable for extracting anthocyanin compounds. Prolonged extraction time can cause a decrease in anthocyanin content, which is associated with anthocyanin degradation by citric acid. The optimal time is 45 minutes with the UAE method.

Keywords: Anthocyanin; Isolation method; Red dragon fruit

INTRODUCTION

Our body cells create ROS (Reactive Oxygen Species), commonly referred to as free radicals, during metabolism. Free radicals are produced by immune cells to destroy viruses and bacteria. [1] Environmental elements like radiation, cigarette smoke, pollution, and pesticides can also create free radicals. The organism generally tolerates free radicals, but oxidative stress results from either an

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overabundance of free radicals or a lack of antioxidants. Environmental elements like radiation, cigarette smoke, pollution, and pesticides can also produce free radicals. The organism generally tolerates free radicals, but oxidative stress results from either an overabundance of free radicals or a lack of antioxidants. Thus, studies on antioxidants derived from natural sources must be carried out.

An inexpensive alternative source of antioxidants that can enhance the quality



of public health is natural materials.^[4] Certain sections of some including the red dragon fruit, naturally produce antioxidants. The plant known as pitaya, or red dragon fruit (Hylocereus polyrhizus), is native to arid tropical areas. One of the most well-liked fruits, it is grown extensively in several tropical nations. In the skin of red dragon fruit. compounds from the betalain. anthocyanin, vitamin E, and vitamin C groups can be found. Red dragon fruit contains anthocyanin chemicals, which can act as antioxidants to protect against free radicals.^[5]

Anthocyanins are natural pigments that give red to purple colors to various parts of plants, including fruits and flowers. Anthocyanins are a type of flavonoid consisting of three carbon atoms bound by an oxygen atom that serves to connect two aromatic benzene rings. (C_6H_6) . In addition to its function as a natural dye, anthocyanins are also known to have strong antioxidant properties. Some benefits anthocyanins are that they are antiinflammatory and anti-cancer, and they protect the cardiovascular system. A compound is said to be a very strong antioxidant if the IC₅₀ value is less than 50, strong (50-100), moderate (100-150), and weak (151-200). The smaller the IC_{50} value, the higher the antioxidant activity.

Many types of solvents can be used to extract anthocyanins, such as water, methanol, or ethanol. Factors such as excessively high extraction temperatures, prolonged extraction times, exposure to light and oxygen, and variations in the pH of the extraction environment can contribute to the degradation of anthocyanin compounds.^[7]

Based on this background, the researchers believe that they need to conduct a literature review to gather literature to understand the factors influencing the isolation of anthocyanin

compounds from dragon fruit peel, such as method factors, solvent factors used, temperature factors, and pH factors. This literature review can serve as a reference for the development of anthocyanin isolation methods in the future.

METHODS

The methods used in this research were obtained from various library and literature sources. The data excavation process was carried out through searches from reliable literature sources such as Google, Google Scholar, and PubMed by for articles on anthocyanin looking isolation and antioxidant tests from red dragon fruit. The keywords used in the search were literature anthocyanin isolation. red dragon fruit. antioxidant. The search data were then further selected within a 10-year range from 2014 to 2024. Several inclusion criteria in the literature search were that the articles were in Indonesian or English, the articles were fully available, and the articles explained the methods of anthocyanin isolation from red dragon fruit. In contrast, the exclusion criteria were literature reviews in the form of articles

RESULTS

Based on the search results from the literature sources, 14 scientific articles were obtained. It was found that the obtained articles used various methods and solvents to isolate anthocyanins, one example being maceration, UAE (Ultrasonic Assisted Extraction), MAE (Microwave Assisted Extraction), and MAHD (Microwave Assisted Hydro Distillation).

Out of the 14 articles used, 10 articles employed maceration as the method for extracting anthocyanins from red dragon fruit peel powder samples.



Table 1. Results of the Review of the Maceration Method of Red Dragon Fruit PeelPowder

Method	Preparation	Instrument	Result	Reference
Maceration	100 g red dragon fruit peel powder	Spectrophotomete	The anthocyanin	[8]
	with aquadest and 10% citric acid	r UV-Vis with $\lambda =$	concentration	
	(1:6) at room temperature for 3 x	520.0 nm	(%) result is	
	24 hour with no stiring		30.4% b/v	
	100 g red dragon fruit peel powder	Spectrophotomete	The anthocyanin	[9] [10]
	with 50 mL of 0.4 M citric acid	r UV-Vis with $\lambda =$	content obtained	
	solvent at a temperature of 20°C	510.0 nm and	was 12.447 mg/L	
	for 2 hours with no stiring	700.0 nm		
	100 g red dragon fruit peel powder	Spectrophotomete	The anthocyanin	[10]
	with 50 mL of 0.4 M citric acid	r UV-Vis with $\lambda =$	content obtained	
	solvent at a temperature of 20°C	510.0 nm and	was 1.225 mg/L	
	for 3 hours with no stiring	700.0 nm		
	10 g red dragon fruit peel powder	Spectrophotomete	The anthocyanin	[11]
	with 30 mL aquadest and 10%	r UV-Vis with $\lambda =$	content obtained	
	citric acid at room temperature for	510.0 nm and	was 4.73 mg/L	
	4 days with 30 second of stirring	700.0 nm		
	once	0 1 1	TTI (1 :	F1.13
	10 g red dragon fruit peel powder	Spectrophotomete	The anthocyanin	[11]
	with 40 mL aquadest and 10%	r UV-Vis with $\lambda =$	content obtained	
	citric acid at room temperature for	510.0 nm and	was 2.40 mg/L	
	4 days with 30 second of stirring	700.0 nm		
	once	0 1 1 1	TDI (I '	[10]
	50 g red dragon fruit peel powder	Spectrophotomete r UV-Vis with $\lambda =$	The anthocyanin content obtained	[12]
	with aquadest and 2% citric acid at	$\kappa = 535.0 \text{ nm}$		
	roomtemperature for 2 x 24 hours with no stiring	333.0 HIII	was 6.38±0.44 mg/50 g	
	Red dragon fruit peel powder with	Spectrophotomete	The maximum	[13]
	ethanol and 10% citric acid (5:1)	r UV-Vis with $\lambda =$	anthocyanin	[13]
	at room temperature for 24 hours	498.6 nm	content was	
	with no replication and no stirring	470.0 IIII	obtained using a	
	with no replication and no staring		10% ethanol and	
			citric acid solvent	
	Red dragon fruit peel powder with	Spectrophotomete	The yield of	[14]
	aquadest and 10% citric acid (1:6)	r UV-Vis with $\lambda =$	anthocyanins was	[1.]
	at room temperature for 3 days	500.0 nm - 550.0	obtained, which	
	with no stirring	nm	is 62.68%	
	50 g red dragon fruit peel powder	Spectrophotomete	The total	[15]
	with aquadest and 2% citric acid	r UV-Vis with $\lambda =$	anthocyanin	. ,
	for 2 x 24 hours at room	535.0 nm	content obtained	
	temperature with no stirring		is 38.33 mg/100	
	-		g	
	20 g red dragon fruit peel powder	Spectrophotomete	The %yield	[16]
	with 96% Ethanol solvent at a	r UV-Vis by	obtained is	
	speed of 15 rpm for 60 minutes at	calculating % yield	0.000275%.	
	room temperature	with $\lambda = 530.0 \text{ nm}$		
	25 g red dragon fruit peel powder	Spectrophotomete	The anthocyanin	[17]
	with 0,5 M citric acid solvent	r UV-Vis with $\lambda =$	content obtained	
	stirred for 150 minutes at room	510.0 nm and	is 11.439 mg/L.	
	temperature	700.0 nm		

Solvents that can be used for extraction include aquadest, nitric acid, and ethanol. The calculation of anthocyanin content obtained from the extraction results is performed using a UV-Vis spectrophotometer.

Out of the 14 articles used, two articles employed ultrasonic-assisted extraction (UAE) as the method for extracting anthocyanins from red dragon fruit peel powder samples. Solvents that can be used for extraction include



aquadest, nitric acid, and ethanol. The anthocyanin calculation of content obtained from the extraction results is performed using a UV-Vis spectrophotometer. Out of the 14 articles used, 1 article employed microwaveassisted extraction (MAE) as the method for extracting anthocyanins from red dragon fruit peel powder samples. Solvents that can be used for extraction include aquadest and nitric acid. The calculation of anthocyanin content obtained from the extraction results is performed using a UV-Vis spectrophotometer.

Out of the 14 articles used, 1 article employed maceration as the method for extracting anthocyanins from red dragon fruit peel powder samples. Solvents that can be used for extraction include aquadest and ethanol. The calculation of anthocyanin content obtained from the extraction results is performed using a UV-Vis spectrophotometer.

Table 2. Results of the Review of the Ultrasonic Assisted Extraction Method of Red Dragon Fruit Peel Powder

Method	Preparation	Instrument	Result	Reference
Ultrasonic Assisted Extraction (UAE)	5 g red dragon fruit peel powder with aquadest and 10% citric acid solvent for 45 minutes and 65% amplitude	UV-Vis with $\lambda = 510.0$	The anthocyanin content obtained is 29.640 mg/L	[18]
	Red dragon fruit peel powder with ethanol, aquadest, and acetic acid (80:19:1) solvent for 15 minutes	Spectrophotometer UV-Vis with $\lambda = 520.0$ nm and 700.0 nm	The anthocyanin content obtained is 0.0241 mg/L	[15]

Table 3. Results of the Review of the Microwave-Assisted Extraction Method of Red Dragon Fruit Peel Powder

Method	Preparation	Instrument	Result	Reference
Microwave	100 g red dragon	Spectrophotometer	The anthocyanin	[19]
Assisted	fruit peel powder	UV-Vis with $\lambda = 535.0$	content obtained was	
Extraction (MAE)	with aquadest and	nm	28.11 mg/100 mg	
	citric acid solvent		extract	
	(4:1) for 6 minutes			

Table 4. Results of the Review of the Microwave Assisted Hydro Distillation Method of Red Dragon Fruit Peel Powder

Method	Preparation	Instrument	Result	Reference
Microwave	100 g red dragon	Spectrophotometer	The anthocyanin	[20]
Assisted Hydro	fruit peel powder	UV-Vis with $\lambda = 510.0$	content obtained	
Distillation	with ethanol and	nm and 700.0 nm	was 52.184 mg/100	
(MAHD)	aquadest solvent		g sample	
	(4:1) for 4 minutes			

DISCUSSION

Based on the search results from the literature sources, 14 scientific articles were obtained. It was found that the obtained articles used various methods and solvents to isolate anthocyanins, one example being maceration, UAE (Ultrasonic Assisted Extraction), MAE

(Microwave Assisted Extraction), and MAHD (Microwave Assisted Hydro Distillation).

A) Maceration Method

Maceration is a method of separating a compound that involves soaking it in an organic solvent at a specific temperature.



The maceration process is considered advantageous for isolating a natural compound due to its low cost and ease of execution. The maceration technique relies on pressure changes inside and outside the cell to break the cell wall and membrane, dissolving secondary metabolites in the cytoplasm. [21]

The amount of solvent used is one of the factors in the amount of compounds extracted in maceration. According to the research results in Table 1, maceration using 30 mL of citric acid and aquadest produced an anthocyanin content of 4.73 mg/L, while maceration using 40 mL of citric acid and aquadest produced an anthocyanin content of 2.40 mg/L. The addition of citric acid gives the solvent acidic properties. Under acidic conditions, plant cell membranes can denature, causing the pigments present in the plants to dissolve more easily into the solvent, making the compounds easier to extract.[13, 22]

Based on the results of the literature study, the more solvent volume used for maceration, the lower the concentration of anthocyanin compounds obtained. from the maceration process. decrease in anthocyanin concentration occurs because the cyanidin groups in the red dragon fruit skin have been extracted by the citric acid solvent, resulting in a lower anthocyanin concentration. so that with the increase in volume, the amount of anthocyanin obtained does increase. This is in accordance with the extraction theory, which states that the concentration of the product decreases with the amount of solvent present.[23]

The acidity level of the solvent also affects the extraction of anthocyanins because anthocyanin compounds are pH-sensitive. In the extraction using citric acid, higher yields were obtained compared to the use of ethanol. This is because anthocyanins are more stable in acidic solutions compared to alkaline or neutral solutions. In acidic solutions, anthocyanins are stable, and in strong

acidic solutions, anthocyanins are very stable, whereas in neutral and basic solutions, anthocyanins become unstable. In a solvent with a high pH, anthocyanins will undergo deprotonation, which weakens the bonds within the molecule, making it more susceptible to degradation. [9]

The duration of the maceration treatment also affects the concentration of anthocyanin compounds extracted during maceration. According to the research results in Table 1, maceration for 2 hours produced an anthocyanin content of 12.447 mg/L. On the other hand, maceration for 3 hours resulted in an anthocyanin content of 1.225 mg/L. The anthocyanin content decreases as the extraction time increases.

Based on the literature review, the longer the extraction time, the higher the amount of anthocyanin compounds obtained. This is because the opportunity for contact between the material and the solvent increases, resulting in a greater vield. However, the increase in extracted compounds will stop when the material reaches saturation point. The decrease in anthocyanin content occurs due to the degradation of anthocyanins, causing the contained anthocyanin levels to decrease. An extraction time that is too long will cause the extract to hydrolyze, while an extraction time that is too short will result in not all active compounds extracted from the material.[14, 18]

B) Ultrasonic Assisted Extraction (UAE) Method

Ultrasonic-assisted extraction (UAE) is a promising extraction method because it yields higher yields and shorter processing times. UAE has the principle of acoustic cavitation, which can damage plant cell walls, allowing bioactive compounds to be released into the environment. UAE has been proven to improve extraction efficiency and extraction time. Additionally, UAE can also be performed at lower temperatures



to avoid damage due to heating.[18, 24]

The use of aquadest and citric acid as solvents in Table 2 has proven to be more effective in extracting anthocyanin compounds, with a yield of 29.640 mg/5 g in the study. In contrast, UAE using ethanol, aquadest, and acetic acid yielded an anthocyanin content of 0.0241 mg/30 Compared to maceration, UAE produced a higher yield, with maceration yielding an anthocyanin content of 38.33 mg/100 g. This is because UAE utilizes ultrasonic waves to enhance penetration of the liquid into the cell walls, resulting in a faster mass transfer rate and, consequently, highe r anthocyanin content.[25]

C) Microwave Assisted Extraction (MAE)

A new method that has received attention in recent times to be developed in the isolation of active compounds, the name of this new method is microwaveassisted extraction (MAE). This method advantages over conventional extraction methods, namely the presence molecular transfer involving microwave energy. The MAE method has the advantage of obtaining extraction yields quickly, as microwave energy causes molecular movement through ion migration and dipole rotation. This very rapid movement generates friction. which ultimately produces thermal energy within the material, causing the cell walls or tissue to break down allowing the solute to be released. [26]

A mixture of aquadest and citric acid, using the MAE method in Table 3, produces an anthocyanin content mg/100 mg, compared 28.11 maceration using the same solvent mixture, which yields an anthocyanin content of 4.73 mg/L. This shows that the MAE method is more effective in extracting anthocyanin compounds compared to maceration because MAE uses microwave waves to generate friction, which ultimately produces heat energy in the material, causing the cell walls or tissue to break down, making it easier for the solvent to attract the compounds.^[27]

The UAE method yields higher results than MAE, with a solvent of aguadest and citric acid yielding 29.640 mg/L, which is not much different from MAE. The UAE method uses a principle almost identical to MAE, which involves using waves to break down cell walls. allowing more compounds to extracted. Based on the literature review, MAE (Microwave-Assisted Extraction) to UAE is stated to be superior (Ultrasound-Assisted Extraction) because MAE is capable of extracting a larger amount of compounds compared to UAE under optimized extraction conditions, even when using a longer extraction time. This is evidenced by the UAE requiring 45 minutes, while MAE only takes 6 minutes.[27]

D) Microwave Assisted Hydro Distillation (MAHD)

Assisted Microwave Hydro Distillation (MAHD) is an advancement Microwave-Assisted Extraction (MAE) that uses electromagnetic waves to heat the solvent. This method is superior because it produces a high extract yield, is easy to use, and is timeefficient. MAHD can be an alternative extraction method for compounds with results similar to those of conventional water distillation. MAHD produces higher anthocyanin content than MAE and maceration, as well as color characteristics that correspond to pH changes.[28]

The MAHD method using ethanol and aquadest solvent (4:1) in Table 4, yields an anthocyanin content of 52.184 mg/100 g. This result is significantly higher compared to maceration, which yields an anthocyanin content of 38.33 mg/100 g. This is because the MAHD method uses microwaves similar to the MAE method to break down the



compound cell walls, thereby increasing the content. The MAHD method is considered more efficient, requiring only 4 minutes, while maceration requires 2 x 24 hours.^[26]

The MAHD method yields higher results compared to the MAE method, which yields 28.11 mg/100 mg. This is because the MAHD method is an improvement of the MAE method with the addition of an integrated distillation process during extraction, resulting in a higher yield.

The results obtained by the UAE method were 29.640 mg/5 g, compared to the MAHD method, where the UAE method yielded a higher concentration. This is because the solvent used in the MAHD method is ethanol. The solvent used in the UAE method is citric acid, which creates an acidic condition in the solvent. This can denature plant cell membranes, making it easier for the pigments in the plants to dissolve into the solvent, thus allowing the compounds to be easily extracted. This results in a lower concentration produced by the MAHD method using ethanol.^[9]

CONCLUSION

Factors that influence the success of the anthocyanin isolation of red dragon fruit include the type and volume of the solvent, the length of time the extraction, and the extraction method used. The ultrasonic assistance method (UAE) is most effective in isolating anthocyanin compounds compared to other methods, with anthocyanin content obtained by 29,640 mg/l. Extraction in acid conditions proved effective in attracting anthocyanin, but the extraction of extraction in acid conditions can cause anthocyanin compounds to degrade. The length of time for anthocyanin extraction in the most optimal acid conditions is 45 minutes with the UAE method. A longer extraction time using an acidic solvent will cause the anthocyanin compounds to degrade and reduce the anthocyanin.

CONFLICT OF INTEREST

There is no conflict of interest in this paper. This article was written independently without affiliation with other parties.

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REFERENCES

- 1. Widayati E. Oxidasi Biologi, Radikal Bebas, dan Antioxidant. Majalah Ilmiah Sultan Agung. 2012; 50(128): 26-32.
- 2. Anisa M, Hardia L, Budiyanto A B. Literature Review: Aktivitas Antioksidan Ekstrak Tongkol Jagung (*Zea mays* L.). JIKES: Jurnal Ilmu Kesehatan. 2023; 2(1): 1-13.
- 3. Nirwana A C, Mutakin M. Review Artikel: Aktivitas Antioksidan dari Suku Rutaceae. Farmaka. 2019; 17(1): 66-76.
- 4. Werdhasari A. Peran Antioksidan Bagi Kesehatan. Jurnal Biotek Medisiana Indonesia. 2014; 3(2): 59-68.
- Winahyu D A, Purnama R C, Setiawati M Y. Uji Aktivitas Antioksidan pada Ekstrak Kulit Buah Naga Merah (Hylocereus polyrhizus) Dengan Metode DPPH. Jurnal Analis Farmasi. 2019; 4(2): 117-121.
- 6. Priska M, Peni N, Carvallo L, Ngapa Y D. Antosianin dan Pemanfaatannya. Cakra Kimia (Indonesian E-Journal of Applied Chemistry). 2018; 6(2): 79-97.
- 7. Augustia V, Wibisono B R, Wara F M R, Kusumaningrum W B. Ekstraksi Antosianin Kulit Buah Naga dan Purifikasi dengan Metode Adsorpsi Menggunakan Amberlite IRC 120. Indonesian Journal of Chemical Research. 2024; 9(1): 96-103



- 8. Nasrullah N, Husain H, Syahrir M. Pengaruh Suhu dan Waktu Pemanasan Terhadap Stabilitas Pigmen Antosianin Ekstrak Asam Sitrat Kulit Buah Naga Merah (Hylocereus polyrizus) dan Aplikasi Pada Bahan Pangan. Chemica. 2020; 21(2): 150-162.
- 9. Ayun Q, Ajeng A. Pengaruh pH Larutan Terhadap Kestabilan Warna Senyawa Antosianin yang Terdapat pada Ekstrak Kulit Buah Naga (*Hylocereus costaricensis*). Jurnal Crystal: Publikasi Penelitian Kimia dan Terapannya. 2023; 4(1): 1-6.
- 10. Ayun Q, Endara R. Optimasi Ekstrak Kulit Buah Naga Merah (*Hylocereus costaricensis*) untuk Mendapatkan Kadar Antosianin Maksimal. Prosiding: Konferensi Nasional Matematika dan IPA Universitas PGRI Banyuwangi. 2022; 2(1): 175-181.
- 11. Widyasanti A, Arsyad M Z, Wulandari E. Ekstraksi Antosianin Kulit Buah Naga Merah (Hylocereus polyrhizus) Menggunakan Metode Maserasi. Jurnal Agroindustri. 2021; 11(2): 72-81.
- 12. Nizori A, Sihombing N. Karakteristik Ekstrak Kulit Buah Naga Merah (Hylocereus polyrhizus) dengan Penambahan Berbagai Kosentrasi Asam Sitrat sebagai Pewarna Alami Makanan. Jurnal Teknologi Industri Pertanian. 2020; 30(2): 228-233
- 13. Meganingtyas W, Alauhdin M. Ekstraksi Antosianin dari Kulit Buah Naga (*Hylocereus costaricensis*) dan Pemanfaatannya sebagai Indikator Alami Titrasi Asam-Basa. AgriTECH. 2021; 41(3): 278-284
- 14. Simanjuntak L, Sinaga C, Fatimah F. Ekstraksi Pigmen Antosianin dari Kulit Buah Naga Merah (*Hylocereus polyrhizus*). 2014; 3(2): 25-29

- 15. Fathurahmi S. Ekstraksi Pewarna Alami Kulit Buah Naga Merah. Jurnal Pengolahan Pangan. 2022; 7(2): 75-79.
- 16. Butar S T B, Rasmito A. Pengaruh Konsentrasi Pelarut dan Lama Ekstraksi Antosianin dari Kulit Buah Naga Merah. Jurnal Riset Teknik. 2023; 2(3): 23-26.
- 17. Rohman M F, Devina S P, Fachri B A, Rahmawati I, Mumtazah Z. Extraction of Anthocyanins from Dragon Fruit Peel Using Solvent Extraction Method. Journal Of Biobased Chemicals. 2024; 4(1): 69-79
- 18. Widyasanti A, Nurlaily N. Wulandari E. Karakteristik Fisikokimia Antosianin Ekstrak Kulit Buah Naga Merah Menggunakan Metode UAE. Jurnal Ilmiah Rekayasa Pertanian dan Biosistem. 2018; 6(1): 27-38.
- Ekstraksi 19. Ingrath W. Pigme n Antosianin dari Kulit Buah Naga Merah (Hylocereus costaricensis) sebagai Pewarna Alami Makanan dengan Menggunakan Microwave (Kajian Waktu Pemanasan dengan Microwave dan Penambahan Rasio Pelarut Aquades dan Asam Sitrat). Jurnal Bioproses Komoditas Tropis. 2015; 3(3): 1-8.
- 20. Shiddiqi Q Y A, Karisma A D. Ekstraksi Senyawa Antosianin dari Kulit Buah Naga Merah (Hylocereus polyrhizus) Menggunakan Metode Microwave Assisted Hydrodistillation (MAHD). Jurnal Chemurgy. 2021; 5(1): 30-37.
- 21. Fakhruzy F. Optimalisasi Metode Maserasi untuk Ekstraksi Tanin Rendemen Tinggi. Menara Ilmu: Jurnal Penelitian dan Kajian Ilmiah. 2020; 14(2): 38-41.
- 22. Ayun Q, Endara R. Optimasi Ekstrak Kulit Buah Naga Merah (*Hylocereus costaricensis*) untuk Mendapatkan Kadar Antosianin



- Maksimal. Prosiding: Konferensi Nasional Matematika dan IPA Universitas PGRI Banyuwangi. 2022; 2(1): 175-181.
- 23. Sahraeni S, Harjanto H, Rahim H. Ekstraksi Antosianin dari Kulit Buah Naga Merah sebagai Pewarna Alami. In Seminar Nasional Hasil Penelitian & Pengabdian Kepada Masyarakat (SNP2M). 2018; 3(1): 105-109
- 24. Suryanto E, Taroreh M R. Ultrasound-Assisted Extraction Antioksidan Serat Pangan dari Tongkol Jagung (*Zea mays* L.). *Chemistry Progress*. 2019; 12(2): 104-110
- 25. Kristina C V M, Yusasrini N A, Yusa N M. Pengaruh Waktu Ekstraksi dengan Menggunakan Metode Ultrasonic Assisted Extraction (UAE) Terhadap Aktivitas Antioksidan Ekstrak Daun Duwet (Syzygium cumini). Jurnal Ilmu dan Teknologi Pangan. 2022: 11(1): 13-21.
- 26. Kamaluddin M H, Lutfi M. Hendrawan Y. Analisa Pengaruh Extraction Microwave Assisted (MAE) Terhadap Ekstraksi Senyawa Antioksidan Catechin pada Daun Teh Hijau (Camellia sinensis) (Kajian Waktu Ekstraksi dan Rasio Bahan: Pelarut). Journal of **Tropical** Agricultural Engineering and Biosystems-Jurnal Keteknikan Pertanian Tropis dan Biosistem. 2014; 2(2): 147-155
- 27. Álvarez R M, Ruíz R A, Barbero G F, Vázquez E M, El-Mansouri F, Brigui J, et al. Comparison between Ultrasound-and Microwave-Assisted Extraction Methods to Determine Phenolic Compounds in Barley (Hordeum vulgare L.). Foods. 2023; 12(14): 2638.
- 28. Lituhayu N, Firmansyah B N, Sativa F I O. Perbandingan Pengambilan Minyak Atsiri Daun Beluntas dengan Menggunakan Metode

Microwave-Assisted Extraction (MAE) dan Microwave Assisted Hydro-Distillation (MAHD). Jurnal Atmosphere. 2024; 5(1): 7-15.