Effects of High g Values on Growth and Chlorophyll Content in Hydrated and Dehydrated Wheat Seeds

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Abstract – Higher g value stimuli (gravitational acceleration more than 1 g and referred as hypergravity) caused by centrifugation have been shown to inhibit elongation growth of various plants. In the present study, effects of high g values were studied on wheat seeds with and without water medium at the time of exposure to high g values. Wheat seeds (variety: LOK-1) were washed with 0.5% fungicide and then 4-5 times with distilled water (D/W). Seeds were then soaked in D/W for 24 hrs. Two different experiments were performed. In initial experiment, soaked seeds were taken into the centrifuge tubes filled with 1 ml D/W. Seeds were exposed to hypergravity ranging from 500 g to 2500 g for 10 minutes. In another experiment, soaked seeds were taken into the centrifuge tubes without water or any other medium and then exposed to high g values. After exposure seeds were immediately sowed on 0.8% agar gel. Results obtained showed that exposure to high g values suppressed growth in wheat seedlings when seeds were exposed to high g values with water as a medium. Chlorophyll content also decreased with increase in g. However, no change in growth and chlorophyll content were observed when seeds were exposed without water medium up to g values as high as 2500. Thus, effects of high g value stimuli depend not only on how much centrifugal force is applied to the seeds but also depends upon how much force is experienced by the inner part of the seeds. Present observation shows that effective centrifugal force experienced by the seeds is different when applied with and without medium.

Key words: Hypergravity, centrifugation, chlorophyll content, wheat

1. Introduction
A high acceleration (g) value, which is also referred as hypergravity, can be produced on earth by centrifugation. Acceleration due to centrifugation has been used to analyze the mechanisms of gravity perception and gravicurvature [1, 2]. Changes in shoot growth have also been studied under hypergravity conditions. Hypergravity stimuli (gravitational acceleration more than 1 g) caused by centrifugation have been shown to inhibit elongation growth of pea epicotyls [3], radish and cucumber hypocotyls [4], cress hypocotyls [5], azuki bean epicotyls [6]. Stem elongation in Arabidopsis thaliana inflorescence stems was suppressed while dry weight of the inflorescence stems increased in hypergravity [7, 8]. Lignin formation and cell wall polysaccharide content in cress hypocotyls [4] and hemicellulosic polysaccharides in wheat coleoptiles [9] increased in hypergravity. Similar results have been obtained for maize coleoptiles and mesocotyls when 3 days old 20-25 mm long maize seedlings were exposed to hypergravity i.e. 300 g for 6 hrs at 25°C in dark [6] and for azuki bean epicotyls when 5 days old 30-35 mm long azuki bean seedlings were exposed to 300 g for 10 hrs at 25°C in dark [10]. In these studies, seedlings were exposed to hypergravity and seedlings were either wrapped with tissue paper or mounted on agar gel at the time of exposure to hypergravity. However, only one report showed decrease in growth and chlorophyll content in rice where seeds were exposed to hypergravity and seeds were exposed to hypergravity with water as a medium inside the centrifuge tubes at the time of hypergravity exposure [11].

Effects of high g values on plants depend upon several factors such as relative centrifugal force (RCF), how long the centrifugal force is applied, shape and size of centrifuge tube, orientation of seeds during centrifugation, properties of medium etc. Depending on seeds orientation at the time of exposure the most affected area can be decided. If the most affected area is the embryo of the seed, suppressed growth will be observed. Properties of medium may also change by centrifugation which could results in difference in growth with and without medium. Here we report the results of effects of hypergravity on wheat seeds with and without water medium at the time of exposure to hypergravity.
2. Materials and Methods

Wheat seeds (variety: LOK-1) were obtained from Sheti Udyog Bhandar, Pune. Seeds were washed with 0.5% fungicide (Uthane M-45) and then 4-5 times with distilled water (D/W). Seeds were then soaked in D/W for 24 hrs. Two different experiments were performed. In initial experiment, eight soaked seeds were taken into each of the five centrifuge tubes filled with 1 ml D/W. Seeds were exposed to hypergravity ranging from 500 g to 2500 g for only 10 minutes. In another experiment, 8 soaked seeds were taken into the centrifuge tubes without water inside the tubes and then exposed to hypergravity ranging from 500 g to 2500 g for only 10 minutes. After hypergravity exposure seeds were immediately sowed on 0.8% agar gel. Seeds not exposed to hypergravity acted as a control.

The experiments were carried out under ambient conditions of temperature (23±4 °C) and humidity (55±10 %) and a light intensity of 15 µmolm⁻²s⁻¹ for 16 hours day⁻¹. Daily growth was observed. Measurements were carried out on the 5th day after sowing. Seed germination percentage of 5 days old seedlings were calculated. Chlorophyll was extracted by using DMF by using the technique given by Porra [12]. Absorption spectra of chlorophyll were taken using a UV-visible spectrometer (Lambda-950, Perkin Elmer, Singapore) and fluorescence spectra at an excitation wavelength of 440 nm using a Luminescence spectrometer (LS-55, Perkin Elmer, Singapore) respectively. Chlorophyll a, chlorophyll b and total chlorophyll contents were calculated by using Arnon’s formula [13]. Experiments were repeated for three times and consistent results were obtained.

3. Results

Figure 1a shows growth of unexposed and exposed wheat seeds to high g values with water as a medium inside the centrifuge tubes at the time of exposure. Results obtained showed that the average seed germination percentage for control, 500 g, 1000 g, 1500 g, 2000 g and 2500 g were 92%, 92%, 71%, 50%, 21% and 21% respectively. No growth was observed in 2000 g while few seeds were germinated even at 2500 g. This value is fluctuating that is sometimes no growth was observed at 2000 g and sometimes at 2500 g. Fig. 1b shows histogram plot of shoot lengths and root lengths for control and hypergravity exposed samples. As seen from the histogram shoot lengths and root lengths were higher in case of control samples as compared with hypergravity exposed samples (p > 0.1, p < 0.025, p < 0.001, p < 0.001 and p < 0.001 for 500 g, 1000 g, 1500 g, 2000 g and 2500 g respectively).
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Figure 1. a) Shows growth of unexposed (control) and seeds exposed to higher ‘g’ values (hypergravity) with water medium. These photos are representative of 3 experiments. b) Histogram showing average shoot lengths and average root lengths for control and hypergravity treated samples.

Figure 2a shows absorption spectra of chlorophyll for control and hypergravity exposed samples extracted from shoots of 5 day old seedlings. Absorption spectra of chlorophyll extracted from shoots show two peaks one at 433.5 nm and another at 669 nm. No shift in wavelength was observed. However, a decrease in the intensity at 433.5 and 669 nm in case of hypergravity treated samples observed suggested that the chlorophyll content has decreased in hypergravity treated samples except at 500 g where slight increase in chlorophyll content was observed (Fig. 2b, Table 1).

Fig. 1b)

Fig. 2a)

Fig. 2b)
<table>
<thead>
<tr>
<th>Chlorophyll content (g/l)</th>
<th>0.0000</th>
<th>0.0005</th>
<th>0.0010</th>
<th>0.0015</th>
<th>0.0020</th>
<th>0.0025</th>
<th>0.0030</th>
<th>0.0035</th>
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**Figure 2.** a) Absorption and b) fluorescence spectra of chlorophyll for unexposed (control) and hypergravity treated samples with water. These spectra are a representative of 3 experiments. c) Chlorophyll content for unexposed (control) and hypergravity treated samples. Average percentage decrease in chlorophyll content in 500 g, 1000 g, 1500 g, 2000 g and 2500 g was 7%, 13%, 43%, 59%, and 60% respectively.

**Figure 3.** Shows growth of unexposed (control) and seeds exposed to higher ‘g’ values (hypergravity) without water inside the centrifuge tubes. These photos are representative of 3 experiments.

Figure 3 shows growth of unexposed and exposed wheat seeds for seeds exposed to hypergravity without water or any other medium inside the centrifuge tubes at the time of exposure. Results obtained showed no change in the average seed germination percentage (100% in all the samples) for control, 500 g, 1000 g, 1500 g, 2000 g and 2500 g. Also, no significant change in the root and shoot lengths and weights observed in control and hypergravity treated samples (Table 1).
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Table 1. Average root and shoot lengths and weights for control and hypergravity treated samples. As shown in figure no significant change in root, shoot lengths and weights was observed.

<table>
<thead>
<tr>
<th>g Value</th>
<th>Average root length (cm)</th>
<th>Average shoot length (cm)</th>
<th>Average Root weight (gm)</th>
<th>Average Shoot weight (gm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>8.27±0.15</td>
<td>9.28±0.33</td>
<td>0.37±0.03</td>
<td>0.42±0.04</td>
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<td>500 g</td>
<td>8.24±0.69</td>
<td>8.87±0.50</td>
<td>0.35±0.04</td>
<td>0.41±0.02</td>
</tr>
<tr>
<td>1000 g</td>
<td>8.41±0.18</td>
<td>8.95±0.67</td>
<td>0.33±0.07</td>
<td>0.40±0.05</td>
</tr>
<tr>
<td>1500 g</td>
<td>8.23±0.10</td>
<td>9.19±0.74</td>
<td>0.35±0.05</td>
<td>0.43±0.04</td>
</tr>
<tr>
<td>2000 g</td>
<td>8.44±0.36</td>
<td>8.67±0.51</td>
<td>0.35±0.06</td>
<td>0.40±0.03</td>
</tr>
<tr>
<td>2500 g</td>
<td>8.40±0.33</td>
<td>9.13±0.77</td>
<td>0.35±0.06</td>
<td>0.42±0.03</td>
</tr>
</tbody>
</table>

Figure 4. a) Absorption and b) fluorescence spectra of chlorophyll for unexposed (control) and hypergravity treated samples. These spectra are a representative of 3 experiments. c) Chlorophyll content for unexposed (control) and hypergravity treated samples. As shown in figure no significant change in chlorophyll content was observed.

As seen from the Fig. 4a and 4b, chlorophyll a, chlorophyll b and total chlorophyll contents were also not changed significantly in hypergravity treated samples as compared to control samples.
4. Discussion

The decrease in seed germination percentage and also decrease in root and shoot lengths for seeds exposed to hypergravity with water as a medium inside the test tubes at the time of exposure suggests that hypergravity suppresses plant growth. This is in agreement to those results obtained for Arabidopsis [7], pea [3], maize [6] and cress [5]. No growth was observed in 2000 g. Some of the seeds germinated even at 2500 g indicated that seeds orientation was not proper at the time of hypergravity exposure. Decrease in chlorophyll content was observed in hypergravity samples. This is in consistent with the results obtained for rice [11] and wheat [14, 15]. However, no change in seed germination percentage, growth and chlorophyll content was observed for seeds exposed to hypergravity without water or any medium inside the test tubes at the time of exposure. These results suggest that effects of hypergravity on growth and chlorophyll content in plants depend on medium through which hypergravity stress.

According to the sedimentation theory, particle in a centrifugal field will experience a centrifugal force defined by:

\[ F_c = m \omega^2 r \]  

Where, \( F_c \) is the centrifugal force, \( m \) is the mass of the particle, \( \omega \) is angular velocity and \( r \) is the distance from the axis.

This force will be opposed by a buoyant force \( (F_b) \) and a frictional force \( (F_f) \). The buoyant force represents the force it takes to displace solvent as the particle moves through the centrifugal field. The frictional force represents the drag on the particle as it passes solvent molecules. These two forces are respectively defined as,

\[ F_b = -m_\sigma \omega^2 r \]  
\[ F_f = -fv \]  

Where, \( m_\sigma \) is the mass of the displaced solution, \( f \) is a frictional coefficient and \( v \) is velocity of the particle.

The particle will move at a velocity such that the total force equals 0, thus

\[ F_c + F_b + F_f = 0 \]  
\[ m \omega^2 r - m_\sigma \omega^2 r - fv = 0 \]  

by replacing \( m_0 \) with \( mV\rho_s \) in equation (3) where \( V \) is partial specific volume of the particle and \( \rho_s \) is density of the solvent, and solving we get

\[ m \omega^2 r = \frac{fv}{1 - V\rho_s} \]  

This equation tells us that, the denser the solution (medium), the higher the centrifugal force experienced by the particle. Thus, the centrifugal force experienced by the seeds was different when seeds were exposed with water medium and without water medium inside the tubes. Further calculation gives net centrifugal force experienced by the seeds with water was approximately 4 times greater than centrifugal force experienced by seeds without water.

5. Conclusion

Effects of hypergravity stimuli depend not only on how much centrifugal force is applied to seeds but also depends upon how much force is experienced by seeds. Centrifugal force experienced by the seeds is different if it is applied through air and the water. Therefore, the growth and chlorophyll content were retarded in hydrated seeds than dehydrated seeds when exposed to higher g values.

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References

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